

## Amaryllidaceae Alkaloids from *Lycoris radiata*

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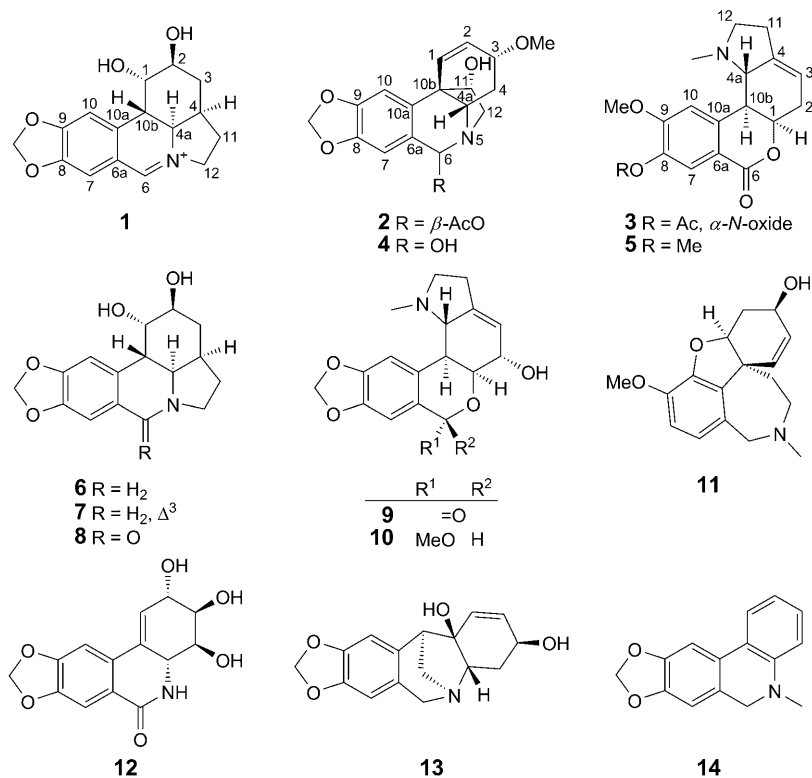
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A phytochemical investigation on bulbs of *Lycoris radiata* resulted in the isolation of three new Amaryllidaceae alkaloids, named 5,6-dehydrodihydrolycorine (**1**), 6 $\beta$ -acetoxycrinamine (**2**), and (+)-8-*O*-acetylhomolycorine  $\alpha$ -*N*-oxide (**3**), together with eleven known alkaloids, **4**–**14**. The structures of the new alkaloids were established by means of spectroscopic methods, and the known compounds were identified by comparison of their data with those in the literature. Compound **2** showed cytotoxicity against HL-60, A-549, and MCF-7 cells, with  $IC_{50}$  values of 8.1, 24.3, and 15.0  $\mu$ M, respectively.

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**Introduction.** – Amaryllidaceae alkaloids have for a long time attracted great interest of synthetic organic chemists because of their several biological activities and their potential diversity in pharmacology [1–9]. So far, more than 100 alkaloids have been isolated from Amaryllidaceae plants [10], which exhibited diverse bioactivities, such as antiviral, insect antifeedant, antineoplastic, and acetylcholinesterase inhibitory activities [10–14]. As part of our search for novel and bioactive compounds, we isolated three new Amaryllidaceae alkaloids, 5,6-dehydrodihydrolycorine (**1**), 6 $\beta$ -acetoxycrinamine (**2**), and (+)-8-*O*-acetylhomolycorine  $\alpha$ -*N*-oxide (**3**), together with eleven known analogs from bulbs of *Lycoris radiata*, a Chinese folk medicine famous for the treatment of poliomyelitis [15]. The new structures were elucidated by means of spectroscopic methods, and the known compounds were identified as 6-hydroxycrinamine (**4**) [16], homolycorine (**5**) [17], dihydrolycorine (**6**) [18], lycorine (**7**) [18], 7-oxodihydrolycorine (**8**) [19], (+)-hippeastrine (**9**) [20], 2 $\alpha$ -hydroxy-6-*O*-methoxylozidine (**10**) [21], galanthamine (**11**) [22], 7-deoxynarciclasine (**12**) [23], pancratinine C (**13**) [24], and 5,6-dihydrobicolorine (**14**) [25]. In addition, compounds **1**–**3** were evaluated for their cytotoxic activities against five human cancer cell lines.

**Results and Discussions.** – Compound **1** was isolated as a yellow powder ( $[\alpha]_D^{25} = +399.7$  ( $c = 0.1$ , MeOH)). The HR-ESI-MS (positive-ion mode) displayed a molecular ion at  $m/z$  288.1228 ( $M^+$ ), corresponding to the molecular formula  $C_{16}H_{18}NO_4$ . The UV absorption bands at 373, 308, 253, and 211 nm suggested an extended chromophore, and a O–CH<sub>2</sub>–O-substituted benzene ring [26], whereas the IR absorption bands at 3405, 3358, 1646, 1605, and 922  $cm^{-1}$  indicated two OH groups and a phenyl function. The <sup>1</sup>H-NMR spectrum (*Table*) displayed two *singlets* for two *para*-positioned aromatic H-atoms ( $\delta(H)$  7.30, 7.16), signal of a OCH<sub>2</sub>O group ( $\delta(H)$  6.18), and a downfield *singlet* corresponding to the H-atom of an iminium salt ( $\delta(H)$  8.86) [27]. The <sup>13</sup>C-NMR spectrum (*Table*) displayed 16 C-atom resonances ascribable to four CH<sub>2</sub> and eight CH



groups (including three  $sp^3$ -C-atoms bearing a heteroatom), and four  $sp^2$  quaternary C-atoms. The above data suggested that compound **1** was an amaryllidaceae alkaloid similar to dihydrolycorine (**6**) [18], except for an imine moiety located between N(5) and C(6) ( $\delta(C)$  163.0) in **1**, as supported by HMCBs of  $\delta(H)$  8.86 (*s*, H–C(6)) with  $\delta(C)$  61.5 (*d*, C(4a)), 139.3 (*s*, C(6a)), 122.3 (*s*, C(10a)), and 57.5 (*t*, C(12)) (*Fig.*). The relative configuration of **1** was elucidated by a ROESY experiment. For biogenetic

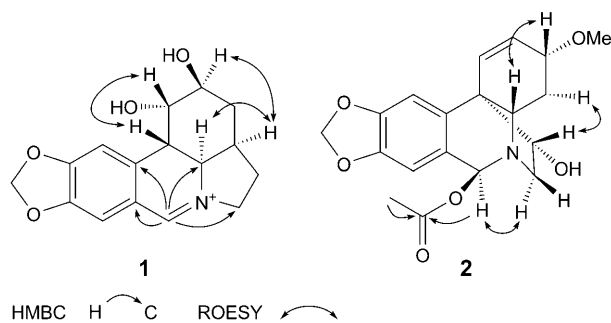


Figure. Key HMBC and ROESY Correlations of **1** and **2**

reasons, H–C(10b) is considered to be  $\beta$ -oriented, and H–C(4a)  $\alpha$ -oriented [18]. Subsequently, the ROESY correlation H–C(10b)/H–C(1) suggested the  $\alpha$ -orientation for the OH group at C(1), and the ROESY correlations of H–C(4a)/H–C(4) and H–C(4)/H–C(2) indicated also  $\alpha$ -orientation for both H–C(4) and H–C(2) (*Fig.*). Finally, detailed analysis of 2D-NMR data established the structure of **1** as 5,6-dehydrodihydrolycorine.

Table. <sup>1</sup>H- and <sup>13</sup>C-NMR Data (500 and 100 MHz, resp.) of **1–3** ( $\delta$  in ppm, *J* in Hz)

Position	<b>1</b> <sup>a)</sup>		<b>2</b> <sup>b)</sup>		<b>3</b> <sup>a)</sup>	
	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)
1	4.48 (br. <i>s</i> )	67.3 ( <i>d</i> )	6.38 ( <i>d</i> , <i>J</i> = 10.1)	126.3 ( <i>d</i> )	4.95 ( <i>d</i> , <i>J</i> = 2.0)	78.0 ( <i>d</i> )
2	3.83 ( <i>ddd</i> , <i>J</i> = 8.5, 4.0, 2.6)	69.1 ( <i>d</i> )	6.41 ( <i>dd</i> , <i>J</i> = 10.1, 1.2)	132.7 ( <i>d</i> )	2.52 ( <i>d</i> , <i>J</i> = 16.8), 2.88–2.91 ( <i>m</i> )	31.5 ( <i>t</i> )
3	1.91 ( <i>dd</i> , <i>J</i> = 11.6, 4.0), 2.14 ( <i>dd</i> , <i>J</i> = 11.6, 2.6)	28.4 ( <i>t</i> )	3.87–3.89 ( <i>m</i> )	72.4 ( <i>d</i> )	5.79 (br. <i>s</i> )	125.9 ( <i>d</i> )
4	2.85–2.88 ( <i>m</i> )	35.1 ( <i>d</i> )	1.99 ( <i>dd</i> , <i>J</i> = 13.3, 5.1), 2.11 ( <i>dd</i> , <i>J</i> = 13.3, 4.5)	27.6 ( <i>t</i> )		141.4 ( <i>s</i> )
4a	4.26–4.28 ( <i>m</i> )	61.5 ( <i>d</i> )	3.63 ( <i>dd</i> , <i>J</i> = 13.3, 4.5)	58.2 ( <i>d</i> )	4.14 ( <i>d</i> , <i>J</i> = 10.0)	79.0 ( <i>d</i> )
6	8.86 ( <i>s</i> )	163.0 ( <i>d</i> )	6.13 ( <i>s</i> )	87.0 ( <i>d</i> )		166.6 ( <i>s</i> )
6a		139.3 ( <i>s</i> )		125.0 ( <i>s</i> )		118.1 ( <i>s</i> )
7	7.30 ( <i>s</i> )	112.6 ( <i>d</i> )	6.65 ( <i>s</i> )	108.8 ( <i>d</i> )	7.15 ( <i>s</i> )	121.3 ( <i>d</i> )
8		149.0 ( <i>s</i> )		146.4 ( <i>s</i> )		143.3 ( <i>s</i> )
9		157.1 ( <i>s</i> )		148.1 ( <i>s</i> )		157.0 ( <i>s</i> )
10	7.16 ( <i>s</i> )	107.1 ( <i>d</i> )	6.83 ( <i>s</i> )	103.0 ( <i>d</i> )	7.52 ( <i>s</i> )	114.1 ( <i>d</i> )
10a		122.3 ( <i>s</i> )		137.1 ( <i>s</i> )		135.4 ( <i>s</i> )
10b	3.09 (br. <i>s</i> )	41.8 ( <i>d</i> )		49.9 ( <i>s</i> )	3.62 ( <i>dd</i> , <i>J</i> = 10.0, 2.0)	38.1 ( <i>d</i> )
11	2.20–2.22 ( <i>m</i> ), 2.28–2.31 ( <i>m</i> )	33.5 ( <i>t</i> )	3.93 ( <i>dd</i> , <i>J</i> = 6.7, 3.0)	78.2 ( <i>d</i> )	2.69–2.72 ( <i>m</i> , 2 H)	26.4 ( <i>t</i> )
12	4.08 ( <i>dt</i> , <i>J</i> = 10.0, 4.4), 4.23 ( <i>t</i> , <i>J</i> = 10.0)	57.5 ( <i>t</i> )	3.27 ( <i>dd</i> , <i>J</i> = 13.5, 3.0), 3.44 ( <i>dd</i> , <i>J</i> = 13.5, 6.7)	58.7 ( <i>t</i> )	3.51 ( <i>t</i> , <i>J</i> = 9.3), 3.70 ( <i>t</i> , <i>J</i> = 9.3)	70.8 ( <i>t</i> )
OCH <sub>2</sub> O	6.18 (br. <i>s</i> )	104.7 ( <i>t</i> )	5.94 (br. <i>s</i> )	101.2 ( <i>t</i> )		
MeN					2.97 ( <i>s</i> )	56.0 ( <i>q</i> )
MeO			3.44 ( <i>s</i> )	56.6 ( <i>q</i> )	3.98 ( <i>s</i> )	57.2 ( <i>q</i> )
MeCO <sub>2</sub>			2.12 ( <i>s</i> )	21.5 ( <i>q</i> )	2.28 ( <i>s</i> )	20.3 ( <i>q</i> )
MeCO <sub>2</sub>				170.4 ( <i>s</i> )		170.2 ( <i>s</i> )

<sup>a)</sup> Recorded in CD<sub>3</sub>OD. <sup>b)</sup> Recorded in CDCl<sub>3</sub>.

Compound **2** was isolated as a colorless oil. The positive-ion-mode HR-ESI-MS displayed an  $[M + H]^+$  peak at  $m/z$  360.1447, corresponding to the molecular formula C<sub>19</sub>H<sub>21</sub>NO<sub>6</sub>. The IR absorption bands at 3386 and 1711 cm<sup>-1</sup> are ascribable to a OH group and an ester CO group, respectively. In the <sup>1</sup>H-NMR spectrum, two *singlets* at  $\delta$ (H) 6.65 (*s*) and 6.83 (*s*) were assigned to two *para*-positioned aromatic H-atoms

H–C(7) and H–C(10), respectively. A broad *singlet* at  $\delta(\text{H})$  5.94 (br. *s*, 2 H) was ascribed to the OCH<sub>2</sub>O H-atoms. Two signals at  $\delta(\text{H})$  6.38 (*d*,  $J=10.1$ ) and 6.41 (*dd*,  $J=10.1, 1.2$ ) were assigned to the olefinic H-atoms H–C(1) and H–C(2), respectively. Two *singlets* at  $\delta(\text{H})$  3.44 (*s*) and 2.12 (*s*) were ascribed to H-atoms of a MeO group and a Me group, respectively. The <sup>13</sup>C-NMR spectrum exhibited 19 C-atom resonances which revealed the existence of a phenyl ( $\delta(\text{C})$  103.0, 108.8, 125.0, 137.1, 146.4, 148.1), an Ac ( $\delta(\text{C})$  21.5, 170.4), a MeO ( $\delta(\text{C})$  56.6), and a OCH<sub>2</sub>O group ( $\delta(\text{C})$  101.2), two O-bearing CH groups ( $\delta(\text{C})$  72.4, 78.2), an sp<sup>3</sup> quaternary C-atom ( $\delta(\text{C})$  49.9), and a downfield sp<sup>3</sup>-CH group ( $\delta(\text{C})$  87.0) bearing two heteroatoms. The above data resembled those of 6-hydroxycrinamine (**4**) [16] except for an additional AcO group ( $\delta(\text{C})$  21.5 (*q*), 170.4 (*s*);  $\delta(\text{H})$  2.12 (*s*, 3 H)) in **2**. The key HMBC of  $\delta(\text{H})$  6.13 (*s*, H–C(6)) with  $\delta(\text{C})$  170.4 (*s*, MeCO<sub>2</sub>) suggested that the AcO group was at C(6) (*Fig.*). The ROESY correlations H–C(4a)/H–C(3) and H–C(11)/H–C(4a) suggested both H–C(3) and H–C(11) to be  $\beta$ -oriented, and the ROESY correlation of H–C(6)/H–C(12 $\alpha$ ) indicated  $\alpha$ -orientation for H–C(6) (*Fig.*). Detailed analysis of 2D-NMR data established compound **2** to be 6 $\beta$ -acetoxyocrinamine.

Compound **3** was obtained as a colorless oil. The IR absorption bands at 1703 and 1657 cm<sup>-1</sup> indicated the existence of CO moieties, while the UV absorption bands at 265 and 212 nm suggested a conjugated moiety. The HR-ESI-MS (positive-ion mode) displayed an  $[M + \text{H}]^+$  peak at  $m/z$  360.1457, 16 mass units higher than that of (+)-8-*O*-acetylhomolycorine [27]. Compound **3** was readily identified as (+)-8-*O*-acetylhomolycorine *N*-oxide from the <sup>1</sup>H- and <sup>13</sup>C-NMR data (*Table*), in particular with the characteristic downfield signals of the C-atom resonances for C(4a) ( $\delta(\text{C})$  79.0), C(12) ( $\delta(\text{C})$  70.8), and MeN ( $\delta(\text{C})$  56.0) with respect to those of (+)-8-*O*-acetylhomolycorine [27]. The ROESY correlation MeN/H–C(4a) suggested  $\alpha$ -orientation of the *N*-oxide [28]. Hence, compound **3** was established as (+)-8-*O*-acetylhomolycorine  $\alpha$ -*N*-oxide.

Compounds **1–3** were evaluated for their cytotoxicity against five human cancer cell lines. Compound **2** showed cytotoxicity against HL-60, A-549, and MCF-7 cells, with *IC*<sub>50</sub> values of 8.1, 24.3, and 15.0  $\mu\text{M}$ , respectively, while the positive control cisplatin gave *IC*<sub>50</sub> values of 2.4, 17.6, and 18.7  $\mu\text{M}$ . Compounds **1** and **3** were inactive (*IC*<sub>50</sub> values > 40  $\mu\text{M}$ ).

### Experimental Part

*General.* Column chromatography (CC): silica gel (SiO<sub>2</sub>, 200–300 mesh; Qingdao Haiyang Chemical Co., Ltd., P. R. China), RP-18 gel (20–45  $\mu\text{m}$ ; Fuji Silysia Chemical Ltd., Japan), and Sephadex LH-20 (Pharmacia Fine Chemical Co., Ltd., Sweden). TLC: GF 254 plates (Qingdao Haiyang Chemical Co., Ltd., P. R. China). Optical rotations: Horiba SEPA-300 polarimeter. UV Spectra: Shimadzu UV-2401A spectrophotometer. IR Spectra: Tenor 27 spectrophotometer with KBr pellets. 1D- and 2D-NMR spectra: a Bruker DRX-500 and an AM-400 spectrometers with TMS as internal standard,  $\delta$  in ppm and  $J$  in Hz. HR-ESI-MS: API-Qstar-Pulsar-1 spectrometer.

*Plant Material.* Bulbs of *L. radiata* were bought from Juhuaacun Chinese Traditional Medicine Market, Kunming, Yunnan Province, P. R. China, and identified by X.-D. L. A voucher specimen has been deposited with the Kunming Institute of Botany, Chinese Academy of Sciences, P. R. China.

*Extraction and Isolation.* The air-dried and powdered sample (15 kg) was powdered and extracted with MeOH at r.t. (3  $\times$  24 h) to give a crude extract (950 g). The extract was partitioned between AcOEt and 0.5% HCl soln. The acidic H<sub>2</sub>O-soluble material was adjusted to pH 9–10 with 10% aq. NH<sub>3</sub> soln. and then extracted with AcOEt to give an alkaloidal extract (68 g). The alkaloidal extract was subjected

to CC (SiO<sub>2</sub>; CHCl<sub>3</sub>/Me<sub>2</sub>CO gradient 1:0, 15:1, 10:1, 5:1, 3:1, 1:1) to give six fractions *Fr. 1–6*. *Fr. 1* (3 g) was subjected to CC (SiO<sub>2</sub>; petroleum ether (PE)/Me<sub>2</sub>CO 15:1 → 10:1) to yield **5** (6 mg) and **14** (80 mg). *Fr. 2* (16 g) was subjected to CC (SiO<sub>2</sub>; PE/Me<sub>2</sub>CO 10:1 → 2:1) to yield **2** (12 mg), **9** (110 mg) and **11** (2 g). *Fr. 3* (7 g) was subjected to CC (SiO<sub>2</sub>; PE/Me<sub>2</sub>CO 5:1 → 1:1), then purified by CC (*RP-18* gel; MeOH/H<sub>2</sub>O 6:4) to yield **4** (130 mg) and **10** (60 mg). *Fr. 4* (9 g) was subjected to CC (*RP-18* gel; MeOH/H<sub>2</sub>O 5:5) to yield **7** (80 mg), **8** (12 mg), and a mixture (2 g). Compound **6** (1.7 g) precipitated from the mixture. *Fr. 5* (11 g) was separated by CC (SiO<sub>2</sub>; CHCl<sub>3</sub>/MeOH 10:1 → 5:1) and further purified by CC (*RP-18* gel; MeOH/H<sub>2</sub>O 4:6) to yield **3** (6 mg), **13** (20 mg), and a mixture (5.8 g). Compound **12** (3.2 g) precipitated from the mixture. *Fr. 6* (7 g) was separated by CC (SiO<sub>2</sub>; CHCl<sub>3</sub>/MeOH 5:1) and further purified by CC (*Sephadex LH-20*; MeOH) to yield **1** (8 mg).

**5,6-Dehydrodihydrolycorine** (= (1*S*,2*S*,3*a**R*,12*b**S*,12*c**R*)-1,2,3,3*a*,4,5,12*b*,12*c*-Octahydro-1,2-dihydroxy[1,3]dioxolo[4,5-*j*]pyrrolo[3,2,1-*de*]phenanthridin-6-ium; **1**). Yellow powder.  $[\alpha]_D^{25} = +399.7$  ( $c = 0.1$ , MeOH). UV (MeOH): 373 (3.95), 308 (3.80), 253 (4.32), 211 (4.73). IR (KBr): 3405, 3358, 1646, 1605, 1589, 1499, 1270, 1033, 922. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Table*. HR-ESI-MS (pos.): 288.1228 ( $M^+$ , C<sub>16</sub>H<sub>18</sub>NO<sub>4</sub><sup>+</sup>; calc. 288.1235).

**6β-Acetoxyocrinamine** (= (3*a*,6*β*,11*S*,13*β*,19*a*)-11-Hydroxy-3-methoxy-1,2-didehydrocrinan-6-yl *Acetate*; **2**). Colorless oil.  $[\alpha]_D^{25} = +22.1$  ( $c = 0.1$ , CHCl<sub>3</sub>). UV (CHCl<sub>3</sub>): 291 (3.79), 240 (3.88). IR (KBr): 3386, 2899, 1711, 1484, 1248, 1060, 932, 868. <sup>1</sup>H- and <sup>13</sup>C-NMR: see *Table*. HR-ESI-MS (pos.): 360.1447 ( $[M + H]^+$ , C<sub>19</sub>H<sub>20</sub>NO<sub>6</sub><sup>+</sup>; calc. 360.1447).

**(+)-8-O-Acetylhomolycorine α-N-oxide** (= (1*R*,5*a**R*,11*b**S*,11*c**S*)-1,2,3,5,5*a*,7,11*b*,11*c*-Octahydro-10-methoxy-1-methyl-1-oxido-7-oxoisochromeno[3,4-*g*]indol-9-yl *Acetate*; **3**). Colorless oil.  $[\alpha]_D^{25} = +113.5$  ( $c = 0.1$ , MeOH). UV (MeOH): 379 (2.67), 307 (3.51), 265 (3.80), 212 (4.61). IR (KBr): 2944, 1703, 1657, 1600, 1451, 1309, 1226, 1066, 909. <sup>1</sup>H- and <sup>13</sup>C-NMR: *Table*. HR-ESI-MS (pos.): 360.1457 ( $[M + H]^+$ , C<sub>19</sub>H<sub>20</sub>NO<sub>6</sub><sup>+</sup>; calc. 360.1447).

**Cytotoxicity Assay.** Five human cancer cell lines (*Sigma*, USA), *i.e.*, breast cancer MCF-7, hepatocellular carcinoma SMMC-7721, human myeloid leukemia HL-60, colon cancer W480, and lung cancer A-549 cells, were used in the cytotoxic assay. Cells were cultured in *RPMI-1640* or in DMEM medium (*Dulbecco's* Modified Eagle Medium; *Hyclone*, USA), supplemented with 10% fetal bovine serum (*Hyclone*, USA) in 5% CO<sub>2</sub> at 37°. The cytotoxicity assay was performed according to the MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2*H*-tetrazolium bromide) method in 96-well microplates [29]. Briefly, 100 μl of adherent cells were seeded into each well of 96-well cell-culture plates and allowed to adhere for 12 h before addition of test compounds, while suspended cells were seeded just before drug addition with initial density of 1 × 10<sup>5</sup> cells/ml. Each tumor cell line was exposed to the test compound dissolved in DMSO at concentrations of 0.0625, 0.32, 1.6, 8, and 40 μM in triplicates for 48 h, with cisplatin (*Sigma*, USA) as a pos. control. After compound treatment, cell viability was detected, and a cell growth curve was graphed. The IC<sub>50</sub> values were calculated by *Reed and Muench's* method [30].

The authors are grateful to the *National Basic Research Program of China* (973 Program 2009CB522300), the *Chinese Academy of Sciences* (KSCX2-YW-R-202), and the *National Natural Science Foundation of China* for partly financial support.

## REFERENCES

- [1] D. H. R. Barton, G. W. Kirby, *J. Chem. Soc.* **1962**, 806.
- [2] N. Hazama, H. Irie, T. Mizutani, T. Shingu, M. Takada, S. Uyeo, A. Yoshitake, *J. Chem. Soc. C* **1968**, 2947.
- [3] Y. Misaka, T. Mizutani, M. Sekido, S. Uyeo, *J. Chem. Soc. C* **1968**, 2954.
- [4] A. G. Schultz, Y. K. Yee, M. H. Berger, *J. Am. Chem. Soc.* **1977**, *99*, 8065.
- [5] S. F. Martin, P. J. Garrison, *J. Org. Chem.* **1982**, *47*, 1513.
- [6] I. H. Sanchez, J. J. Soria, F. J. Lopez, M. I. Larraza, H. J. Flores, *J. Org. Chem.* **1984**, *49*, 157.
- [7] K. A. Parker, H. J. Kim, *J. Org. Chem.* **1992**, *57*, 752.
- [8] Y. Kita, M. Arisawa, M. Gyoten, M. Nakajima, R. Hamada, H. Tohma, T. Takada, *J. Org. Chem.* **1998**, *63*, 6625.

- [9] C.-A. Fan, Y.-Q. Tu, Z.-L. Song, E. Zhang, L. Shi, M. Wang, B. Wang, S.-Y. Zhang, *Org. Lett.* **2004**, *6*, 4691.
- [10] A. Kornienko, A. Evidente, *Chem. Rev.* **2008**, *108*, 1982.
- [11] C. Griffin, N. Sharda, D. Sood, J. Nair, J. McNulty, S. Pandey, *Cancer Cell Int.* **2007**, *7*, 10.
- [12] X.-S. Liu, J. Jiang, X.-Y. Jiao, Y.-E. Wu, J.-H. Lin, Y.-M. Cai, *Cancer Lett.* **2009**, *274*, 16.
- [13] O. Arrigoni, R. Arrigoni-Liso, G. Calabrese, *Nature* **1975**, *256*, 513.
- [14] O. Arrigoni, R. Arrigoni-Liso, G. Calabrese, *Science* **1976**, *194*, 332.
- [15] Y. Tsiang, P.-Y. Li, 'Flora of China', Science Press, Beijing, 1977, Vol. 63, p. 18.
- [16] Y. Tsuda, N. Kashiwaba, V. Kumar, *Chem. Pharm. Bull.* **1984**, *32*, 3023.
- [17] R. Suau, A. I. Gómez, R. Rico, M. P. Vázquez Tato, L. Castedo, R. Riguera, *Phytochemistry* **1988**, *27*, 3285.
- [18] A. Evidente, M. R. Cicala, I. Giudicianni, G. Randazzo, R. Riccio, *Phytochemistry* **1983**, *22*, 581.
- [19] Y. Tsuda, T. Sano, J. Taga, K. Isobe, J. Toda, S. Takagi, M. Yamaki, M. Murata, H. Irie, H. Tanaka, *J. Chem. Soc., Perkin Trans. I* **1979**, 1358.
- [20] M. Kihara, K. Konishi, L. Xu, S. Kobayashi, *Chem. Pharm. Bull.* **1991**, *39*, 1849.
- [21] G. R. Almanza, J. M. Fernández, E. W. T. Wakori, F. Viladomat, C. Codina, J. Bastida, *Phytochemistry* **1996**, *43*, 1375.
- [22] R. Vlahova, D. Krikorian, G. Spassov, M. Chinova, I. Vlahov, S. Parushev, G. Snatzke, L. Ernst, K. Kieslich, W.-R. Abraham, W. S. Sheldrick, *Tetrahedron* **1989**, *45*, 3329.
- [23] G. R. Pettit, N. Melody, *J. Nat. Prod.* **2005**, *68*, 207.
- [24] J. C. Cedrón, J. C. Oberti, A. Estévez-Braun, Á. G. Ravelo, M. Del Arco-Aguilar, M. López, *J. Nat. Prod.* **2009**, *72*, 112.
- [25] F. Viladomat, J. Bastida, G. Tribo, C. Codina, M. Rubiralta, *Phytochemistry* **1990**, *29*, 1307.
- [26] M. P. V. Tato, L. Castedo, R. Riguera, *Heterocycles* **1988**, *27*, 2833.
- [27] J. Bastida, C. Codina, F. Viladomat, M. Rubiralta, J.-C. Quirion, B. Weniger, *J. Nat. Prod.* **1992**, *55*, 122.
- [28] F.-W. Lin, P.-L. Wu, T.-S. Wu, *Chem. Pharm. Bull.* **2001**, *49*, 1292.
- [29] T. Mosmann, *J. Immunol. Methods* **1983**, *65*, 55.
- [30] L. J. Reed, H. Muench, *Am. J. Hygiene* **1938**, *27*, 493.

Received May 1, 2010